In the Specification:

Please replace the sequence listing filed on October 11, 2006 with the attached sequence listing.

Please amend the specification on page 23 lines 9-13 as follows:

Sequence of *Lin ext ba*:
5'-GAT GCC GGC CAC GAT GCG TCC GGC-3' (SEQ ID NO. 1)

Sequence of *Hae sub fo*:
5'-C GTC ATG **GFC** TAT GCG GGC GAC CAC ACC CGT CCT GTG GAT-3' (SEQ ID NO. 2)

Please amend the specification on page 27 carrying over onto page 28 as follows:

For the expression of the proteins and the production of the covalent protein-DNA complexes the samples were incubated at 30 °C for 150 min. Subsequently, the aqueous phase containing the DNA-protein fusions was extracted from the emulsion as follows:
The samples were centrifuged for 10 min. at ~~7.000~~ 7,000 rpm, whereafter the water compartments sedimented at the bottom of the reaction vials. The supernatant (oil phase) was suctioned off and 150 µl buffer were added (buffer consisting of: TBS (Tris-buffered saline) with 1 mM $CaCl_2$ (=TBSC), pH 7.4, 5 µM biotinylated double-stranded DNA fragments for blocking the magnetic beads employed later on [5'-biotin-GGA GCT TCT GCA TTC TGT GTG

CTG-3' (SEQ ID NO. 3) (Qiagen)], 1 µM competing double-stranded DNA fragments [5'-ATC TAA **GGC** CAA TGT ACT AGA C**GG CC**A TTC CAG ATG CA**G GCC** AAG CGT ACA TAC **GGC** CTA GCT AAA TCA A**GG CC**G TAT CGT-3' (SEQ ID NO. 4), substrate sequence for *M.Hae III* in bold letters (Qiagen)]) followed by 1 ml diethyl ether. Subsequently, the sample was shaken with a vortex for 2 x 10 sec. After the separation of the water phase and the oil phase the aqueous phase lying below was removed with a pipette and dried in a 24-microtiter plate for 10 min., so that the remaining diethyl ether was allowed to evaporate completely.

During the extraction of the aqueous phase 25 µl magnetic beads coated with streptavidine (Dynabeads, Dynal, Norway) were incubated with biotinylated, Calmodulin-binding peptide (400 nM, biotin-CAAARWKKAFIAVSAANRFKKIS (SEQ ID NO. 5) (Montigiani et al., 1996) or with biotinylated anti-Flag antibody M2 (2 µl/50 µl beads, M2 antibody, Sigma-Aldrich) for 15 min. The Calmodulin-binding peptide was used to select the *M.Hae III*-Calmoduilin-DNA fusions located in the aqueous phase of the emulsion, whereas the anti-Flag antibody was employed as a negative control. After the incubation of the magnetic beads with peptides or antibodies these were washed once with TBSC 0.1% Tween 20 (Fluka). Subsequently the beads were blocked for 15 min. at room temperature with biotinylated DNA fragments (5 µM) [5'-biotin-GGA GCT TCT GCA TTC TGT GTG CTG-3' (SEQ ID NO. 3) (Qiagen)].

Please amend the specification on the bottom of page 29

carrying over onto page 30 as follows:

To the transcription/translation mixture a mixture of 10^9 DNA molecules in total was added, wherein a factor of 4200 more DNA molecules coded for the fusion protein *M.Hae III*-ED-B than for *M.Hae III*-Calmodulin. The selection experiment was done with magnetic beads that had been coated either with Calmodulin-binding peptides or with anti-Flag antibodies (M2, Sigma-Aldrich). The result of the experiment was evaluated by real-time PCR. However, the magnetic beads were not used for the real-time PCR directly, but the selected DNA molecules were first amplified in a PCT with the primers Ampl ba (5'-CCC GCG AAA TTA ATA CGA CTC A-3', (SEQ ID NO. 6) Qiagen) and Ampl fo (5'-AAA ACC CCT CAA GAC CCG TT-3', (SEQ ID NO. 7) Qiagen). The PRC was performed with the following temperature program:

94 °C (3 min.) → [94 °C (45 sec.) → 51 °C (1 min.) → 72 °C (100 sec.)]$_{35\ cycles}$ → 72 °C (3 min.) → 4 °C.